A New Holopyga from the Western United States (Hymenoptera: Chrysididae).

By Wm. G. Bodenstein, Cornell University, Ithaca, New York.

In a recent loan of Chrysididae from the United States National Museum there is a large series of the following new species of *Holopyga*.

Holopyga taylori*, new species.

Head: In anterior aspect transverse, oval; in dorsal aspect slightly wider and about as long as the pronotum. Vertex with shallow, well-spaced punctures becoming very fine and widely spaced on the occiput. Facial basin with a moderate vertical depression, polished, and with traces of transverse striations; punctured at the sides similarly to the vertex. Clypeus elevated between the antennae, with a few very fine punctures. Mandibles with three teeth. Antennae with the second joint of the

flagellum slightly longer than the fourth.

Thorax: Prothorax punctured similarly to the occiput, the fine punctures interspersed with large, shallow, almost obsolete punctures; the punctuation not much heavier at the sides than on the disc. Mesothorax with very fine, well-separated punctures, becoming close and dense at the anterior ends of the parapsidal sutures; the punctures becoming sparser toward the posterior end of the segment. Mesopleurae with the punctuation indistinct and with traces of fine striations. Scutellum smooth and polished with some traces of obsolete punctures. Postscutellum reticulately punctured on the disc with a small, oblique, rectangular area on each side which is transversely striated. Propodeum reticulately punctured; oblique foveolae roughly triangular in shape with the broad base of the triangle at right angles to the lateral angles of the propodeum; about in the middle of the base of the triangle there is a low, blunt projection. Between the foveolae is a rugose, almost striate area below which there is a vertical ridge. Lateral angles of the propodeum short and acute. Tegulae dark brown, smooth and polished. Wings with the discoidal cells completely lacking; not infuscated. Claws with two inner teeth.

Abdomen: In dorsal aspect the abdomen is egg-shaped, about three-quarters as wide as long; all segments are of about equal length on the median line. First segment with fine

^{*}I name this species for Dr. Leland H. Taylor whose notes and material on the Chrysididae have been of inestimable value to me.

punctures and fine transverse striations, impunctate in the center of the anterior border. Second and third segments punctured as the first, the punctures becoming a little closer together posteriorly. Third abdominal segment truncate at the apex, the truncate portion roughly about one-third the width of the segment at the base. Apical margin very thin, not membranous and not turned under.

Color: Blackish-bronze in color with blue and green reflections on sides of thorax and abdomen. Tibiae and tarsi light

brown.

Female: The female differs from the male as follows: vertex and occiput evenly puctured with fine punctures, slightly larger than those of the male. Facial basin with the transverse striations more distinct. Thorax more evenly punctured; mesopleurae with more distinct punctures. Scutellum more distinctly punctured at the sides. Punctures of the propodeum continued evenly between the oblique foveolae. Abdomen not as heavily punctured, the transverse striations very faint. Face metallic blueish-green in color.

Length: 2-3.5 mm.

Туре. — &; Hollister, Twin Falls County, Ірано. August 7, 1930. (Plot 3a.) [United States National Museum.] Allotype.— ♀. The same locality, August 21, 1930. (Outside.) [United States National Museum.]

Paratypes. — Seventeen males, twenty females, as follows: IDAHO: 4\$; 1\$; Hollister, Twin Falls County; June 13 to July 24, 1931; (D. E. Fox; on Sophia sophia and Salsola pestifera); 2\$; August 13, 1929; 5\$; August 7, 1930; 1\$; July 30, 1929; (F. B. Hinnenkamp; on Salsola parviflora). 2\$; Adelaide, July 25, 1928; (on Sophia sophia); and 1\$; September 14, 1929; on (Salsola pestifera); and 1\$; July 21, 1927; (on Salsola filipes). 2\$; Burley, June 16, 1931; also August 7, 1929; (on Salsola pestifera, at edge of cultivated area). 1\$; Castelford, June 28, 1928; (on Sophia sophia); and 1\$; August 25, 1927; (on Atriplex rosea). 1\$; Berger; September 5, 1930; (on Salsola pestifera). 1\$; Kimama; May 15, 1931; (F. B. Hinnenkamp; on Norta altissima). [All U. S. N. M.]

Missouri: 39; St. Louis; January 8, 1920; (Phil Rau;

numbers 3812 and 3813); [U. S. N. M.]

CALIFORNIA: 19 San Diego County; April; (Coquillett);

[U. S. N. M.]. 63; 39; Emeryville, October 26 and November 3, 1938; (J. W. MacSwain).

This species may be easily distinguished from the previously described North American species of *Holopyga* by its small size, its reduced punctuation, the absence of the discoidal cells, the two teeth in the tarsal claw, and by the apical margin of the third abdominal segment which is very thin and not turned under. A further discussion of this and a related species will be made in a revisionary paper now in preparation.

A Treatment for Crumpled Wings of Odonata Nymphs to Disclose Their Venation.

By May K. Gyger, Ithaca, New York.

Dragonfly nymphs may often be determined to the genus by means of the venation in the wing pads. However, in the latter part of the ultimate nymphal instar the wings are much crumpled and their characters thereby obscured. In the course of some work on nymphs from the Philippine Islands a very peculiar and unique specimen with wing pads badly crumpled was found and Dr. Needham suggested to the author to see whether anything could be done to stretch the wings so that the venation could be seen. The following method is the outcome of much trial and error and, while probably not the best possible, does give usable results.

To prepare an anisopterous wing pad for study, clip it from the nymph and remove the outer layer by cutting along its entire hind margin with a razor blade, then tease the wing out with a dissecting needle. Immerse the wing in boiling water for about three minutes and next place it in twenty drops of a one percent solution of potassium hydroxide. The membrane will begin to expand in a few seconds. Leave it in this solution until it has ceased swelling. Remove, place in boiling water again and float therefrom onto an ordinary glass slide. If the specimen has been in alcohol for a long time further treatment is necessary. Holding the slide in both hands with the fingers underneath and the thumbs on top, gently pull and